

Antihypertensive activity of flowering twigs of *Calotropis procera* (Ait.) is predominantly mediated through vasorelaxant pathway

Wahib Ullah Khan¹, Niaz Ali^{1,2}, Abid Ullah³, Muhammad Nabi¹

ABSTRACT

Objective: To evaluate the crude methanolic (Cp.Cr) and aqueous (Cp.Aq) extracts of *Calotropis procera* Aiton for acute toxicity and antihypertensive effects, validating its ethnomedicinal use in cardiovascular disorders.

Methods: Phytochemical screening was conducted to determine the chemical constituents of *C. procera*. Acute toxicity was assessed in BALB/C mice using oral doses of 10, 100, 1000, 1500, and 2000 mg/kg. Vasorelaxant effects were evaluated using rabbit aortic strips pre-contracted with 1 μ M norepinephrine and 80 mM potassium chloride (high K⁺) to explore the mechanism of action. Antihypertensive activity was assessed in hypertensive Sprague-Dawley rats at doses of 1 and 10 mg/kg orally, using the tail-cuff method to measure systolic blood pressure.

Results: Phytochemical analysis revealed the presence of flavonoids, saponins, cardiac glycosides, and terpenoids. The extract was non-toxic up to 1500 mg/kg, while a 75% mortality rate was observed at 2000 mg/kg. Both Cp.Cr and Cp.Aq induced dose-dependent relaxation in K⁺ and norepinephrine-induced contractions. The EC₅₀ for Cp.Aq was 0.15 mg/ml for KCl-induced and 0.13 mg/ml for NE-induced contractions. A significant reduction in systolic blood pressure (p<0.05) was observed, particularly with the aqueous fraction of *C. procera*.

Conclusion: The flowering twigs of *C. procera* exhibit significant antihypertensive effects, likely mediated through vasorelaxation via inhibition of voltage-gated calcium channels. These findings support its potential use in hypertension management and provide evidence of its safety and pharmacological efficacy.

Keywords: Calotropis (MeSH); Antihypertensive Agent (MeSH); Hypertension (MeSH); Toxicity Tests, Acute (MeSH); Vasodilation (MeSH).

THIS ARTICLE MAY BE CITED AS: Khan WU, Ali N, Ullah A, Nabi M. Antihypertensive activity of flowering twigs of *Calotropis procera* (Ait.) is predominantly mediated through vasorelaxant pathway. Khyber Med Univ J 2025; 17(2): 149-56. <https://doi.org/10.35845/kmuj.2025.23835>

INTRODUCTION

Calotropis procera is an evergreen perennial shrub belonging to the family Apocynaceae. It typically thrives in dry and xerophytic conditions. The plant exudes a white gluey latex from its stem or leaves upon incision or cuts to the aerial parts. Its flowers are fleshy and range in color from white to pinkish, clustered at the ends of twigs (Figure 1).¹ *C. procera* is native to Africa but is also widely distributed in Asia, where it is commonly known as "Calotropis," "dead sea fruit," and "giant milkweed." In Pakistan, it is referred to as "Aak".² Traditional uses of *C. procera* include remedies for rheumatism, eczema, jaundice, fever, cold, and diarrhoea,

with various parts of the plant being used medicinally. The roots are known to treat ailments such as leprosy, cough, asthma, diarrhoea, eczema, and dysentery, while the latex alleviates joint pain and swelling. Additionally, the flowers are employed in managing asthma, loss of appetite, stomach problems, and cough, and they serve as a general tonic. The root bark is used to address intestinal worms, skin disorders, and ascites.¹ Phytochemical studies reveal that *C. procera* contains numerous bioactive compounds, including alkaloids, steroids, terpenoids, tannins, phenols, flavonoids, and sugars.⁴ The flowers of *C. procera* have demonstrated notable anthelmintic properties in sheep,⁵ and their pharmacological activities include

- 1: Department of Pharmacology, Institute of Basic Medical Sciences (IBMS), Khyber Medical University, Peshawar, Pakistan
- 2: Department of Pharmacology, College of Medicine, Shaqra University, Shaqra 11961, Saudi Arabia
- 3: Department of Pharmacy, Shaheed Benazir Bhutto University Sheringal, Dir Upper, Khyber Pakhtunkhwa (KP), Pakistan

Email: niazali@su.edu.sa

Contact #: +92-333-9791763

Date Submitted: October 26, 2024

Date Revised: February 14, 2025

Date Accepted: February 20, 2025

analgesic, antipyretic, anti-inflammatory, and antimicrobial effects.⁶ Hepatoprotective properties of the flowers have also been reported.⁷ Furthermore, the latex exhibits antifungal, antibacterial, and antioxidant activities.⁸ The roots possess antifertility activity in female rats.⁹ Despite extensive documentation of these pharmacological properties, the potential antihypertensive effects of *C. procera* remain underexplored.

Hypertension is a prevalent global health challenge, contributing significantly to cardiovascular morbidity and mortality. Although traditional knowledge associates *C. procera* with antihypertensive benefits,¹⁰ comprehensive scientific validation of this effect, particularly for flowering twigs, is lacking. Existing studies predominantly focus on the plant's antimicrobial, anti-inflammatory, and hepatoprotective properties, leaving a critical gap in understanding its cardiovascular effects. Addressing this gap is essential to harnessing its potential for therapeutic applications in managing hypertension.

This study aims to investigate the antihypertensive activity of the flowering twigs of *C. procera* (Ait.) through in vivo screening. Additionally, acute toxicity assessments will be conducted to establish a safe dosage range. All experiments were conducted in the Pharmacology Laboratory of Institute of Basic Medical Sciences,



Figure 1: Flowering twigs (A) and aerial parts of *C. procera* (B).

Khyber Medical University. Animals were housed in the animal facility of Khyber Medical University, and in vivo experiments were performed within the laboratory to minimize disturbances.

METHODS

Plant materials and preparation of extract:

Fresh flowering twigs were collected from Bannu region of Khyber Pakhtunkhwa, Pakistan. Professor Hashim Khan, Department of Botany, Government Post Graduate College Bannu identified the plant specimen. The flowering twigs were air-dried in the shade at room temperature for 25 days. Once dried, 2.0 kg of the plant material was finely ground using a standard grinder, yielding 1.8 kg of powdered material. This powdered sample was then macerated with 80% hydro-methanol at room temperature for seven days. Following the maceration process, the mixture was filtered using standard-grade filter paper. The resulting filtrates were concentrated under reduced pressure using a rotary evaporator, producing 150 g of a semi-solid crude methanolic extract (designated as Cp.Cr). The solubility of the extract was subsequently tested in normal saline and distilled water.

Phytochemical analysis and isolation of abundant phytochemicals:

Standardized protocols were employed to identify various classes of phytochemicals including alkaloids, terpenes, sterols, flavonoids, saponins, cardiac glycosides, and tannins. For fractionation of abundant phytochemicals, a series of

organic solvents were employed. First, 150g of Cp.Cr was suspended in distilled water. Then the dissolved materials along with water were successively fractionated in a higher order of polar solvents, yielding *n*-hexane, ethyl acetate, chloroform, *n*-butanol, and residual aqueous portions respectively.

Drugs and standards: Analytical-grade chemicals were used in experiments. Norepinephrine was bought from Merck, Germany. Acetylcholine was purchased from BDH, England. Acetylcholine and norepinephrine were purchased from England. Test solutions and suspensions were prepared on the same day of experiments in the pharmacology laboratory.

Experimental animals: BALB/C mice weighing between 25 and 40 g were procured from RAS Traders, Lahore. Male Sprague Dawley rats, weighing between 150 and 250 g, were also obtained from the same supplier. Additionally, rabbits were purchased from the local market in Peshawar, Khyber Pakhtunkhwa, Pakistan. All the animals were maintained in the animal house in their respective cages. Advanced Study & Research Board and Ethical Board of the Khyber Medical University approved the study protocols (Approval No. Dir/KMU-EB/AA/000419). The research was carried out according to the globally accepted guidelines for the use of laboratory animals and care as found in European Community guidelines (EEC Directive of 1986).

Data recording: Lab chart 7 was used for recording changes in isometric tension using a Force Transducer

(Model No: MLT 0225 Pan Lab S.I) coupled with a bridge amplifier connected with 4 channels Power lab (Model No: 4/25T). Antihypertensive activity was recorded using a non-invasive blood pressure (NIBP) controller system supplied with 4 channels Power Lab (Model No: 4/25T).

Physiological solutions used in the experiments:

Three types of Krebs solutions were used in the experiments. Normal Krebs solution contains (mM) NaCl 118.2, KCl 4.7, KH₂PO₄ 1.3, MgSO₄ 1.2, NaHCO₃ 25.0, Glucose 11.7, and CaCl₂ 2.5. While the composition of K-Normal Calcium free Krebs solution was: K-Normal (Ca++ free) Krebs solution with ingredients in (mM): NaCl 118.2, KCl 4.7, KH₂PO₄ 1.3, MgSO₄ 1.2, NaHCO₃ 25.0, Glucose 11.7, and ethylenediamine tetraacetic acid (EDTA) 0.1. The concentration of Krich (Calcium-free) Krebs solution was: (NaCl 50.58, KCl 50, KH₂PO₄ 1.26, MgSO₄ 3.10, NaHCO₃ 23.8, Glucose 11.1, and EDTA) 0.1. Fresh solutions were used in the experiments.

Stock solutions used in the experiments:

Stock solutions of *C. procera* were prepared at concentrations of 10, 100, 150, and 200 mg/ml for acute toxicity evaluation. For the assessment of vasorelaxation effects, stock solutions with concentrations of 3, 30, and 300 mg/ml were prepared and used in the experiments. Isolated aortae were maintained in Krebs normal solution for tissue studies. The observed vasorelaxant effects were further translated into in vivo experiments using rats. Additionally, stock solutions of 1 mg/ml and 10 mg/ml were prepared for antihypertensive activity and subsequent dosing.

Acute toxicity studies: Acute toxicity studies of Cp.Cr were conducted using BALB/C mice. The animals were divided into four groups, each consisting of four mice. One group was denoted as negative control group which was administered with normal saline (10 mL/kg), while three groups were denoted as treated groups which were given the extract at doses 10, 100 and 1000 mg/kg orally. Signs and symptoms of toxicity and mortality were observed over a 24-hour follow-up period,

following our previously reported methodology.¹² Further, the extract was checked in test doses 1500 and 2000 mg/kg for mortality and morbidity. The LD50 value was calculated according to our reported procedure.^{12,13}

Determination of vasorelaxation effects:

The test samples were evaluated for their potential vasorelaxation effects using rabbit aortic strip preparations. Briefly describing, rabbits were slaughtered. Their thoracic aorta was isolated and maintained in Krebs solution continuously aerated with carbogen gas. The aortae were carefully cleared of connective tissues where rings measuring 2-3 mm in thickness were prepared. These segments were maintained in Krebs solution with the following concentrations (in mM): NaCl 118.2, KCl 4.7, MgSO₄ 1.2, KH₂PO₄ 1.3, Glucose 11.7, NaHCO₃ 25.0, CaCl₂ 2.5 with pH 7.4. The preparations were mounted in a 10 mL organ bath maintained at 37°C and continuously aerated with carbogen gas (95% oxygen and 5% carbon dioxide). One end of each aortic strip was attached to a stainless-steel triangle connected to a force-displacement transducer, while the other end was secured to the organ bath to stabilize the isolated tissue. An initial tension of 1 g was applied to the aortic strips and the preparations were allowed to stabilize for 1 hour. Force Transducer (Model No: MLT 0225 Pan Lab S.I) coupled with bridge amplifier connected with 4 channels Power lab (Model No: 4/25T) was used for measuring changes in isometric tension.¹⁴

The aorta was considered intact with functional endothelium if it exhibited relaxation greater than 80% of its control response upon the application of acetylcholine. Aortae were deemed denuded if they failed to show significant relaxation to acetylcholine following the denuding procedure.¹⁵ Test concentrations of the samples (0.001, 0.01, 0.1, 1, and 10 mg/ml) were applied cumulatively at 2-minute intervals to KCl-induced and norepinephrine-induced contractions.

Effects of test samples on norepinephrine-induced contractions in aortae: We employed a similar experimental protocol as previously described. To investigate the

effect of Cp.Cr on receptor-operated calcium channels, we assessed its ability to inhibit contractions induced by 1 μ M norepinephrine in denuded aortic rings. The rationale behind this approach is that the suppression of norepinephrine-induced contractions in aortic tissue provides evidence for the blockade of calcium influx through these channels.^{16,17} As endothelial cells are rich with α 1 adrenergic receptors, activation of these receptors leads to vasoconstriction through the influx of calcium through receptors-operated calcium channels. Therefore, in calcium-free Krebs solution, the NE (1 μ M) induced contractions follow the stimulation of α 1 adrenergic receptors that facilitate calcium release from internal stores.^{17,18} It is important to note that vascular smooth muscle relaxation does not always correlate with the inhibition of voltage-gated calcium channels. Therefore, to confirm the involvement of voltage-gated calcium channels, we tested the Cp.Cr in similar concentrations of 0.001, 0.01, 0.1, 1, and 10 mg/ml on 80 mM KCl-induced contractions.¹⁶ Mean Effective Concentrations (EC₅₀) were recorded for test samples and amlodipine.^{19,20} The experiments were run four times.

Responses were recorded using Lab Chart 7 which was supplied by Power Lab, AD Instruments Australia.

Antihypertensive activity: As preliminary isolated studies in aortae showed vasorelaxation on contractions induced by 80 mM KCl and on 1 μ M Norepinephrine induced contractions, we sought to further investigate its potential via in vivo effects by assessing its impact on the systolic blood pressure of the experimental animals. Non-invasive blood pressure (NIBP) controller system supplied with 4 channels Power Lab (Model No: 4/25T), was used for determining the antihypertensive effect of test samples and amlodipine, a standard CCB. Depomedrol® was used for the induction of hypertension through the subcutaneous route as per our reported procedure.^{21, 22} Prior to experimentation, rats underwent a 7-10-day acclimation period, including 30 minutes of daily restraint training. The animals were then randomly divided into three groups (n=5/group). The first two groups received oral

administrations of Cp.Cr and the aqueous vehicle Cp.Aq, respectively, at doses of 1 and 10 mg/kg. The tail cuff method was used for measuring systolic blood pressure as described.²³ Lab chart 7 was used for recording data using (NIBP) apparatus. Mean systolic blood pressure for treated groups and control hypertensive groups were recorded. Amlodipine in a test dose of 0.0714 mg/kg was used as a standard antihypertensive drug.²¹

Statistical analysis: Response based on isometric tension was plotted as % of control on y-axis versus its respective test concentrations that were plotted on x-axis. Means EC50 were calculated using Graph Pad Prism version 8. Systolic blood pressure (mmHg) was plotted on y-axis versus its respective doses plotted on x-axis. Student 't' test was used to compare the mean of test concentrations with mean of control at 95% CI with 'P' value less or equal to 0.05.

RESULTS

Preliminary phytochemical

composition: Cp.Cr was yielded with a quantity of 150 g. Subsequent fractionation resulted in the following yields: ethyl acetate (7.4 g), n-hexane (6.5 g), n-butanol (3.1 g), chloroform (2.06 g), and an aqueous fraction (100g). Phytochemical screening revealed the presence of flavonoids, saponins, cardiac glycosides, and terpenoids. Alkaloids and tannins were absent. The presence of flavonoids and cardiac glycosides was detected at moderate levels. Notably, a strong positive reaction for saponins was observed, with the highest concentration found in the aqueous fraction (100g).

Toxicity profile: Acute oral toxicity studies indicated no mortality at doses up to 1500 mg/kg b.w. in mice. However, a 75% mortality rate was observed with a dose of 2000 mg/kg. The LD50 of the extracts is below 2000 mg/kg.

Effects of Cp.Cr, Cp.Aq on KCl-induced contractions: The potency of the Cp.Cr is quantified by its EC₅₀ value, which is the concentration required to produce 50% of the maximum effect. Effects of Cp.Cr, Cp.Aq and amlodipine on potassium chloride and norepinephrine-induced contractions

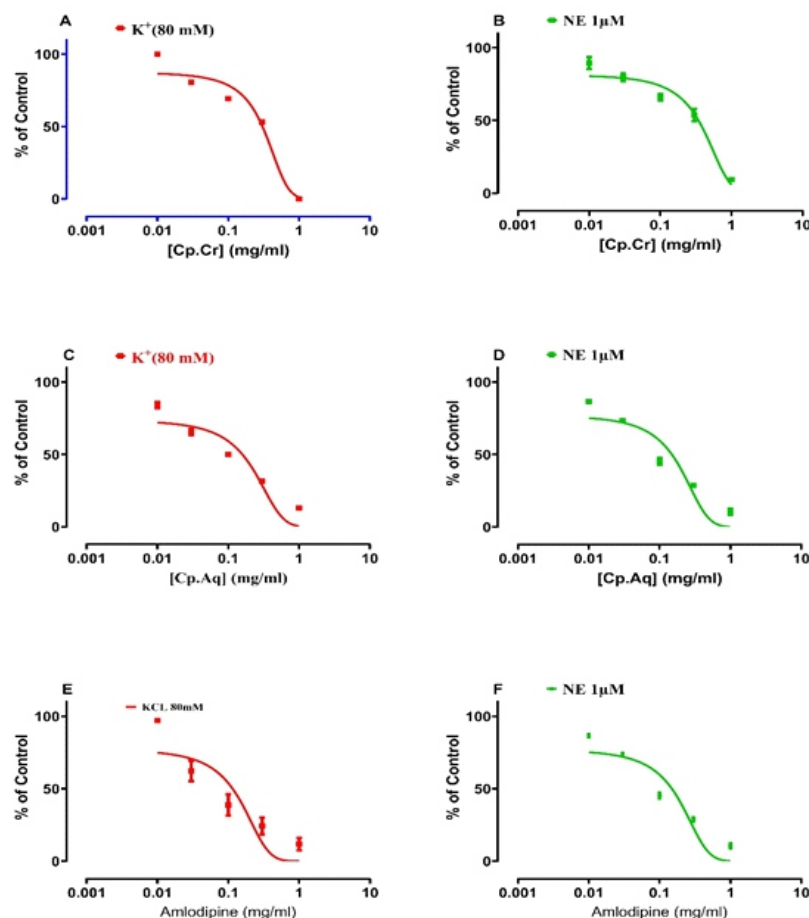


Figure 2: Effects of Cp.Cr, Cp.Aq and amlodipine on contractions induced by potassium chloride (KCl) and norepinephrine (NE) in isolated aortic strips preparations. All values are represented as mean \pm SD (n=4).

on rabbits' thoracic aortae are shown in Figure 2. The crude extract demonstrated inhibitory effects on contractions induced by 80 mM KCl (Figure 2A) starting at 0.03 mg/ml and relaxed at 1 mg/ml with mean EC_{50} value of 0.31 ± 0.06 mg/ml (Table I). The crude extract also completely relaxed norepinephrine (NE $1 \mu\text{M}$) induced contractions (Figure 2B) at 1 mg/ml while its relaxing effect started at 0.01 mg/ml. A relatively potent and significant effect of Cp.Aq was observed on KCl-induced contractions and $1 \mu\text{M}$ NE-induced contractions with their respective EC_{50} as 0.15 ± 0.04 and 0.13 ± 0.03 mg/ml. The EC_{50} of Amlodipine on KCl-induced and $1 \mu\text{M}$ NE-induced contractions were almost superimposable with that of EC_{50} of Cp.Aq fraction.

Effect on systolic blood pressure: Both the extracts Cp.Cr and Cp.Aq

exhibited a dose-dependent hypotensive effect. At a dose of 10 mg/kg, both Cp.Cr and Cp.Aq portion significantly reduced systolic blood pressure ($P < 0.05$) as illustrated in Figure 3. The mean values for these effects are presented in Table II.

DISCUSSION

The present study demonstrated that the crude (Cp.Cr) and aqueous (Cp.Aq) extracts of *Calotropis procera* possess significant vasorelaxant and hypotensive effects. These findings were supported by in vitro vascular relaxation against KCl- and norepinephrine-induced contractions, as well as dose-dependent reductions in systolic blood pressure in hypertensive rats, with Cp.Aq showing potency comparable to amlodipine.

Hypertension, a major global health challenge, significantly increases the risk of cardiovascular diseases. While

synthetic antihypertensive drugs such as calcium channel blockers and beta-blockers are widely used, their side effects and cost necessitate the search for safer, affordable alternatives from natural sources. In this context, traditional medicine offers a rich resource, with numerous plants historically employed for managing hypertension. Among them, *Calotropis procera* is a medicinal plant of significant ethnobotanical relevance in treating cardiovascular ailments piqued our interest, prompting a scientific investigation into its antihypertensive potential. This study focuses on evaluating the vasorelaxant and antihypertensive effects of *C. procera* using both methanolic and aqueous extracts, selected for their ability to cover a broad polarity spectrum and efficiently extract diverse phytochemicals.²⁰⁻²² Alongside, a toxicity profile was established to ensure its safety for potential therapeutic use. A foundational step in developing any phytotherapeutic agent is a comprehensive safety assessment. In our study, toxicity tests of the crude methanolic extract of *C. procera* indicated a wide safety margin.²²⁻²⁵

Animals tolerated doses up to 1500 mg/kg without mortality, achieving 100% survival. However, administering a dose of 2000 mg/kg led to significant mortality (75%), reflecting a steep increase in toxicity beyond the established threshold. Observed adverse effects included respiratory depression and sedation, with mild symptoms at lower doses (10 and 100 mg/kg) escalating to severe respiratory depression at higher doses (1000 and 1500 mg/kg). These findings highlight a therapeutic window, warranting further acute and subacute toxicity studies to optimize dosage and better understand the mechanisms of adverse effects. The vasorelaxant effects of *C. procera* were explored using isolated aortic rings pre-contracted with norepinephrine (NE) and potassium chloride (KCl) (Figure 2, Table I). Both the crude methanolic extract (Cp.Cr) and aqueous extract (Cp.Aq) induced significant relaxation, providing mechanistic insights into their antihypertensive action. Relaxation of NE-induced contractions suggests inhibition of receptor-operated calcium channels (ROCCs), while relaxation of contractions induced by high-

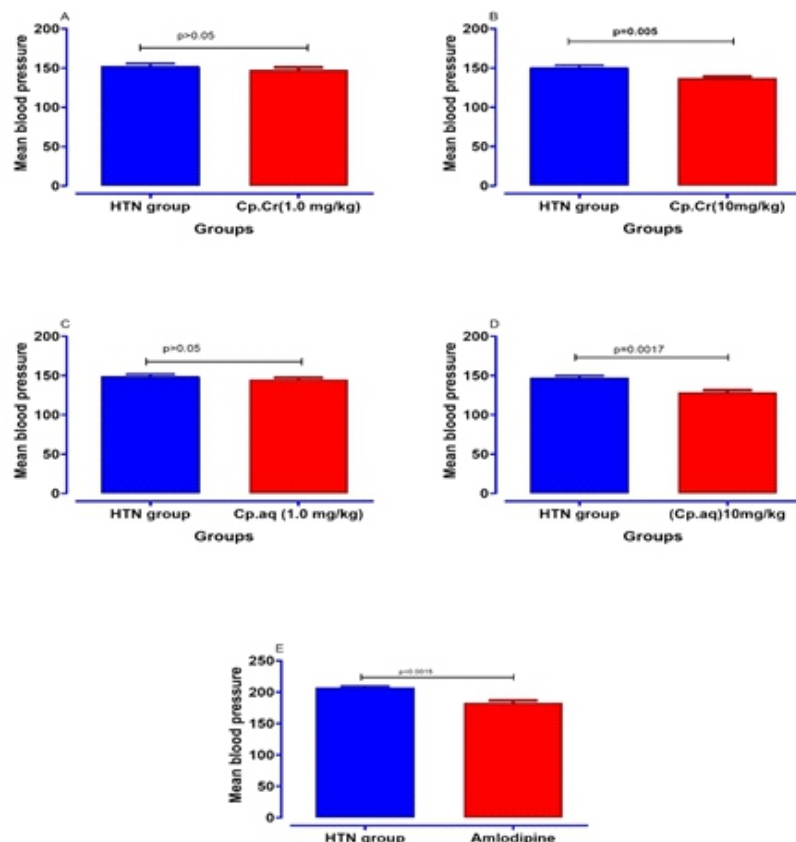


Figure 3: All the values are represented as mean \pm standard deviation (n=4). BP: Blood pressure; Cp.Cr: crude extract of flowering twigs of *C. procera*; Cp.Aq: Aqueous fraction of flowering twigs of *C. procera*.

Table I: Mean EC_{50} values of Cp.Cr and Cp.Aq in isolated denuded aortae of rabbits' preparations

Variable	Simple					
	Cp.Cr		Cp.Aq		Amlodipine	
Types of contractions	80 mM KCl-induced	1 μ M NE-induced	80 mM KCl-induced	1 μ M NE-induced	80 mM KCl-induced	1 μ M NE-induced
Mean ($EC_{50} \pm SD$) (mg/ml)	0.31 \pm 0.06	0.14 \pm 0.02	0.15 \pm 0.04	0.13 \pm 0.03	0.12 \pm 0.03	0.13 \pm 0.03

All the values are represented as mean \pm standard deviation (n=4). NE: norepinephrine; KCl: potassium chloride; Cp.Cr: crude extract of flowering twigs of *C. procera*; Cp.Aq: Aqueous fraction of flowering twigs of *C. procera*.

concentration KCl (80 mM) indicates inhibition of voltage-gated calcium channels (VGCCs), specifically L-type channels.²⁶⁻²⁸ Compounds that relax KCl (>30 mM)-induced contractions are generally recognized as VGCC inhibitors, supporting the hypothesis that calcium influx modulation underlies the vasodilatory effect of *C. procera*. Additionally, the potential role of

endothelium-mediated mechanisms was considered. Nitric oxide (NO), a key mediator in vascular tone regulation, activates guanylyl cyclase, increasing cyclic guanosine monophosphate (cGMP) levels and promoting smooth muscle relaxation. However, the relaxation response in endothelium-denuded tissues points to an endothelium-independent pathway,

reinforcing calcium channel blockade as a primary mechanism.^{29,30} The aqueous extract (rich in saponins) exhibited superior activity compared to the crude extract (Figure 3, Table II), suggesting the presence of concentrated active vasorelaxant constituents in the aqueous fraction. The in vivo translation of these findings demonstrated a dose-dependent reduction in both systolic and diastolic blood pressure. The aqueous extract displayed more potent antihypertensive effects than the methanolic extract, achieving a reduction in systolic blood pressure equivalent to 75% of the response elicited by amlodipine, a standard calcium channel blocker. This aligns with in vitro results, confirming the aqueous extract's potential to inhibit both VGCCs and ROCCs, leading to effective vascular relaxation. Overall, *C. procera* holds significant promise as a source of novel antihypertensive agents. However, comprehensive pharmacological profiling and advanced toxicological studies are warranted to advance its development into a safe and effective therapeutic option.

CONCLUSION

The present study demonstrates that *Calotropis procera* is safe for oral administration at doses up to 1500 mg/kg, confirming its potential for therapeutic use with minimal toxicity at this dosage. The aqueous extract of *C. procera*, which is rich in saponins, exhibits a significant and dose-dependent blood pressure-lowering effect. This hypotensive action appears to be mediated through a vasorelaxation mechanism, primarily involving the inhibition of voltage-gated calcium channels, thereby reducing calcium influx into vascular smooth muscle cells. These findings highlight the plant's potential as a natural antihypertensive agent and provide a foundation for future research into its bioactive compounds and their mechanisms of action. Further studies are warranted to explore its long-term safety, efficacy in clinical settings, and potential interactions with conventional antihypertensive therapies."

ACKNOWLEDGMENTS

The authors extend their sincere gratitude to Khyber Medical University

Table II: The effect of crude Cp.Cr, Cp.Aq and amlodipine on systolic blood pressure in hypertensive

Dose (PO)	Baseline Blood Pressure (mm Hg)	After treatment Blood Pressure (mm Hg)	Fall in BP (mm Hg)	Fall in BP as % of Amlodipine
Cp.Cr 1 mg/kg	152.42 ± 8.04	147.8 ± 7.2	4.6 ± 4.20	19.2
Cp.Cr 10 mg/kg	150.8 ± 6.3	137 ± 5	13.8 ± 8.40	57.2
Cp.Aq 1 mg/kg	149.2 ± 5.8	145.2 ± 4.5	4 ± 1.4	16.7
Cp.Aq 10 mg/kg	147.2 ± 6.1	128.6 ± 6.5	18.6 ± 4.7	75
Amlodipine 0.0714 mg/kg	207.5 ± 4.2	183 ± 7.8	24 ± 3.5	100

All the values are represented as mean ± standard deviation (n=4). BP: Blood pressure; Cp.Cr: crude extract of flowering twigs of *C. procera*; Cp.Aq: Aqueous fraction of flowering twigs of *C. procera*.

for providing institutional support and facilitating the conduct of this research.

The authors would like to thank the Deanship of Scientific Research at Shaqra University for supporting this work."

The authors further acknowledge Mr. Waqar Ali and Mr. Hamid Shams for their significant contributions to manuscript drafting and for their technical assistance in tabulating EC₅₀ values using GraphPad Prism. Their efforts greatly enriched the quality and precision of this work.

REFERENCES

1. Khairnar A, Bhamare S, Bhamare H. *Calotropis procera*: an ethnopharmacological update. *Adv Res Pharm Bio* 2012;2(2):142-56.
2. Ranjit PM, Eswara Rao G, Krishnapriya M, Nagalakshmi V, Silpa P, Anjali M. An overview of phytochemical and pharmacological activities of *Calotropis procera*. *FS J Pharm Res* 2012;1(2):18-25.
3. Dwivedi A, Chaturvedi M, Gupta A, Argal A. Medicinal utility of *Calotropis procera* (Ait.) R. Br. as used by natives of village Sanwer of Indore District, Madhya Pradesh. *Int J Pharm Life Sci* 2010;1(3):188-90.
4. Paul A, Kumar A. Review on pharmacological properties of aaka (*Calotropis procera*). *Int J Econ Plants* 2018;5(3):157-62.
5. Iqbal Z, Lateef M, Jabbar A, M-

hammad G, Khan MN. Anthelmintic activity of *Calotropis procera* (Ait.) Ait. F. flowers in sheep. *J Ethnopharmacol* 2005;102(2):256-61. <https://doi.org/10.1016/j.jep.2005.06.022>

6. Mascolo N, Sharma R, Jain S, C-apasso F. Et-hnopharmacology of *Calotropis procera* flowers. *J Ethnopharmacol* 1988;22(2):211-21. [https://doi.org/10.1016/0378-8741\(88\)90129-8](https://doi.org/10.1016/0378-8741(88)90129-8)
7. Ramachandra Setty S, Quereshi AA, Viswanath Swamy A, Patil T, Prakash T, Prabhu K, et al. Hepatoprotective activity of *Calotropis procera* flowers against paracetamol-induced hepatic injury in rats. *Fitoterapia* 2007;78(78):451-4. <https://doi.org/10.1016/j.fitote.2006.11.022>
8. Ahmed M, Khan R, Shahzaib S, Khan A, Zaif A, Ahmed W. Antifungal, antioxidant and antibacterial activities of *Calotropis procera*. *Int J Biosci* 2014;5:75-80.
9. Kamath J, Rana A. Pharmacological activities of ethanolic extract of *Calotropis procera* roots. *Indian Drugs* 2003;40(5):292-5.
10. Atawodi SE-O, Olowoniyi OD, D-aikwo MA. Ethnobotanical survey of some plants used for the management of hypertension in the Igala speaking area of Kogi State, Nigeria. *Ann Res Rev Biol* 2014;4535-43.

<https://doi.org/10.9734/ARRB/2014/8633>

11. Evans WC. Trease and Evans' Pharmacognosy E-Book: Elsevier Health Sciences; 2009. ISBN: 9780702048838
12. Ali N, Sultana U, Shah SWA, Nabi M, Shah I, Ahmed G. Acute toxicity and analgesic activity of crude flavonoids of *Achillea Wilhelmsii* and *Teucrium Stocksianum*. *Khyber Med Univ J* 2016;8(1):7-11.
13. Ali N, Ahmed G, Shah SWA, Shah I, Ghias M, Khan I. Acute toxicity, brine shrimp cytotoxicity and relaxant activity of fruits of *Callistemon citrinus curtis*. *BMC Complement Altern Med* 2011;11(1):99. <https://doi.org/10.1186/1472-6882-11-99>
14. Cheng Y-W, Kang J-J. Mechanism of vasorelaxation of thoracic aorta caused by xanthone. *Eur J Pharmacol* 1997;336(1):23-8. [https://doi.org/10.1016/S0014-2999\(97\)01224-7](https://doi.org/10.1016/S0014-2999(97)01224-7)
15. Li J, Li W-Q, Yao Y. Vasorelaxation Effect of Estrone Derivate EA204 in Rabbit Aorta. *Scientifica* 2016;2016:7405797. <https://doi.org/10.1155/2016/7405797>
16. Karaki H, Ozaki H, Hori M, Mitsui-Saito M, Amano K-I, Harada K-I, et al. Calcium movements, distribution, and functions in smooth muscle. *Pharmacol Rev* 1997;49(2):157-230.
17. Musha S, Watanabe M, Ishida Y, Kato S, Konishi M, Tomoda A. A phenoxazine compound, 2-amino-4, 4α-dihydro-4α-7-dimethyl-3H-phenoxazine-3-one reverses the phenylephrine or high-K⁺ induced contraction of smooth muscles in rat aorta and guinea pig tenia cecum. *Biol Pharm Bull* 2005;28(8):1521-3. <https://doi.org/10.1248/bpb.28.1521>
18. Cauvin C, Saida K, van Breemen C. Extracellular Ca²⁺ dependence and diltiazem inhibition of contraction in rabbit conduit arteries and mesenteric resistance

- vessels. Blood Vessels 1984; 21(1): 23-31. <https://doi.org/10.1248/bpb.28.1521>
19. Ali N, Begum R, Faisal MS, Khan A, Nabi M, Shehzadi G, et al. Current statins show calcium channel blocking activity through voltage gated channels. BMC Pharmacol Toxicol 2016;17(1):43. <https://doi.org/10.1186/s40360-016-0086-5>
20. Shahid A, Khan DA, Aati HY, Sherif AE, Ovatlarnporn C, Hussain M, et al. Chemical profiling and biological activities of *Dipterygium glaucum* Decne.: an in-vivo, in-vitro and in-silico evaluation. South Afr J Bot 2023; 160: 715-30. <https://doi.org/10.1016/j.sajb.2023.07.033>
21. Rao H, Ahmad S, Aati HY, Basit A, Ahmad I, Ghalloo BA, et al. Phytochemical screening, biological evaluation, and molecular docking studies of aerial parts of *Trigonella hamosa* (branched Fenugreek). Arab J Chem 2023;16(7):104795. <https://doi.org/10.1016/j.arabjc.2023.104795>
22. Lei M, Wang L, Olatunde OO, Singh S, Ovatlarnporn C, Basit A, et al. UPLC-ESI-QTOF-MS profiling, antioxidant, antidiabetic, antibacterial, anti-inflammatory, antiproliferative activities and in silico molecular docking analysis of *Barleria strigosa*. Chem Biol Technol Agric 2023;10(1):73. <https://doi.org/10.1186/s40538-023-00451-2>
23. Basit A, Ahmad S, Sherif AE, A-ati H-Y, Ovatlarnporn C, Khan MA, et al. New mechanistic insights on *Justicia vahlii* Roth: UPLC-Q-TOF-MS and GC-MS based metabolomics, in-vivo, in-silico toxicological, antioxidant based anti-inflammatory and enzyme inhibition evaluation. Arab J Chem 2022; 15(10): 104135. <https://doi.org/10.1016/j.arabjc.2022.104135>
24. Basit A, Ahmad S, Ovatlarnporn C, Arshad MA, Saleem MF, Khurshid U, et al. Unrivalled insight into possible biopharmaceutical application of *Justicia Vahlia* Roth. (acanthaceae): chemodiversity, in vitro bioactivities, and computational analysis. Chem Biodiversity 2024;21(12):e202401432. <https://doi.org/10.1002/cbdv.202401432>
25. Gul H, Hussain A, Javaid F, Khan KU, Basit A, Arafat M, et al. Novel insights into the anti-asthmatic effect of *Raphanus sativus* L. (Raphani Semen): targeting immune cells, inflammatory pathways and oxidative stress markers. J Ethnopharmacol 2024;325:117851. <https://doi.org/10.1016/j.jep.2024.117851>
26. Gilani AH, Khan A-u, Jabeen Q, Subhan F, Ghafar R. Antispasmodic and blood pressure lowering effects of *Valeriana wallichii* are mediated through K⁺ channel activation. J Ethnopharmacol 2005; 100(3): 347-52. <https://doi.org/10.1016/j.jep.2005.05.010>
27. Ali N, Ali W, Ullah A, Ahmad S, Al-saiari AA, Al-mehmadi M, et al. Atorvastatin and Fluvastatin potentiate blood pressure lowering effect of Amlodipine through vasorelaxant phenomenon. Medicina 2023;59(6):1023. <https://doi.org/10.3390/medicina59061023>
28. Ali W, Ali N, Ullah A, Rahman SU, Ahmad S. Pitavastatin and Lovastatin exhibit calcium channel blocking activity which potentiate vasorelaxant effects of amlodipine: a new futuristic dimension in statin's pleiotropy. Medicina 2023; 59(10): 1805. <https://doi.org/10.3390/medicina59101805>
29. Lorenz JN. A practical guide to evaluating cardiovascular, renal, and pulmonary function in mice. Am J Physiol Regul Integr Comp Physiol 2002;282(6):R1565-R82. <https://doi.org/10.1152/ajpregu.00759.2001>
30. Chan SS-K, Choi AO-K, Jones RL, Lin G. Mechanisms underlying the vasorelaxing effects of butyridenephthalide, an active constituent of *Ligusticum chuanxiong*, in rat isolated aorta. Eur J Pharmacol 2006;537(1):111-7. <https://doi.org/10.1016/j.ejphar.2006.03.015>

AUTHORS' CONTRIBUTION

The following authors have made substantial contributions to the manuscript as under:

WUK & MN: Acquisition, analysis and interpretation of data, drafting the manuscript, approval of the final version to be published

NA: Conception and study design, drafting the manuscript, approval of the final version to be published

AU: Conception and study design, critical review, approval of the final version to be published

Authors agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

CONFLICT OF INTEREST

Authors declared no conflict of interest, whether financial or otherwise, that could influence the integrity, objectivity, or validity of their research work.

GRANT SUPPORT AND FINANCIAL DISCLOSURE

Authors declared no specific grant for this research from any funding agency in the public, commercial or non-profit sectors

DATA SHARING STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request

DISCLAIMER

The abstract of the work was published in the abstract book of the 5th International Meeting on Complementary and Alternative Medicine & Therapies, September 18-19, 2017 Charlotte, USA in Curr Res Integr Med (<https://www.pulsus.com/conference-abstracts-files/antihypertensive-activity-of-flowering-twigs-of-calotropis-procera.pdf>)



This is an Open Access article distributed under the terms of the [Creative Commons Attribution 4.0 International License](https://creativecommons.org/licenses/by/4.0/).

KMUJ web address: www.kmu.edu.pk

Email address: kmu@kmu.edu.pk